Improvement Efficacy of Influenza Nanovaccine in Combination with Hemokinin-1 Molecular Adjuvant

Atefeh Dehghan 1, Shahla Shahsavandi 2*, and Leila Jabalameli 1

1. Department of Microbiology, Karaj Branch, Islamic Azad University, Karaj, Iran
2. Razi Vaccine and Serum Research Institute, Agricultural Research Education and Extension Organization, Karaj, Iran

Abstract

**Background:** H9N2 avian influenza viruses have the potential to become the next human pandemic threat and next generation vaccine technologies are needed. Current studies introduce nanoparticles as a proper vaccine delivery vehicle for induction of protective immunity. In this study, the efficacy of chitosan nanoparticle-based H9N2 influenza vaccine with and without hemokinin-1 (HK-1) as a molecular adjuvant to induce protective immunity against the virus was examined.

**Methods:** The H9N2 antigen was prepared in MDCK cells and inactivated with formalin. The inactivated antigen alone and in combination with HK-1 was encapsulated into chitosan nanoparticles. Groups of BALB/c mice received chitosan nanoparticle-based H9N2 antigen alone or in combination with HK-1 in a prime/boost platform via eye drop method. To evaluate the efficacy of the adjuvanted-nanovaccine candidate, systemic antibody responses were compared among the groups of animals.

**Results:** Serological analysis indicated that mice receiving the HK-1/H9N2 nanoparticle formulation induced higher antibody titers that were sustained until the end of experiment. However, in the immunized mice, influenza specific antibody titers were comparable to that in the animals which were immunized either with inactivated antigen alone or the H9N2 nanoparticles without HK-1 adjuvant.

**Conclusion:** The data demonstrate the synergy between HK-1 as an adjuvant and chitosan nanoparticles as a delivery antigen/adjuvant carrier in the improvement of influenza immune responses.

**Keywords:** Chitosan, Immunization, Influenza vaccines, Influenza virus, Nanoparticles

Introduction

Influenza is a worldwide contagious respiratory disease of human, mammals and birds. For decades, chemoprophylaxis and vaccination have been used against influenza infection. Current influenza vaccination program uses either attenuated vaccines or inactivated vaccines to induce protective immunity. Despite the availability of influenza vaccines since the 1930s, direct transmission of avian virus subtypes to human and emergence of new antigenic variants made the efficacy of these vaccines ambiguous. Candidate H9N2 subtype as a potential next pandemic agent has raised a concern about new controlling strategies.

As a primary idea, development and applications of recombinant proteins and DNA vaccines increased attention. Several studies revealed that vaccines based on the virus-coding DNA vaccines including Hemagglutinin (HA), nucleoprotein, matrix, and neuraminidase antigens are able to increase the level of CD8+ Cytotoxic T Lymphocyte (CTL) and generate antigen-specific immune responses. These vaccines as well as traditionally inactivated vaccine fail to induce protective immunity when administered without adjuvant because of their poor immunogenic features. Thus, to enhance immune response, influenza antigen is formulated commonly with one of the licensed adjuvants such as aluminum salts, oil-in-water emulsions, and liposomes.

To induce stronger responses as well as CTL-mediated immunity, molecular adjuvants are also characterized. Compared with the current adjuvants, they act better by enhanced antigen persistence, increased antigen uptake, and recruitment of Antigen Presenting Cells (APCs). Moreover, the inactivated vaccines usually administered via intramuscular injection may fail to reach the APCs and induction of immune responses. To overcome the difficulty, antigen delivery system is considered as one crucial mechanism in targeting the APCs for increasing the efficacy of vac-
vaccines. Currently, many antigen and adjuvant delivery carriers can facilitate the induction of immunity.\textsuperscript{18-20}

Protection against influenza virus infection greatly depends on both humoral and mucosal immune responses to a major surface glycoprotein, HA.\textsuperscript{21} Secretory IgA antibodies are the main mediator of upper respiratory tract adaptive mucosal immunity against the influenza. Induction of the immunity is thought to be improved by mucosal delivered vaccine due to a correlation between the amount of IgA antibodies and the efficacy of protection against the infection.\textsuperscript{18,19} Therefore, a potent influenza vaccine should induce and elevate the antibodies in the host respiratory tract when administrated intranasal. The protection rate is dependent on antigen delivery system that can effectively elicit immune responses.

Nanoparticles are served recently for mucosal antigen delivery system for a wide range of vaccines. The uptake of these particles by APCs can be activated and it can trigger the proliferation of B-cells resulting in class switching of antigen specific plasma cells to produce IgA.\textsuperscript{20} Among all the available nanoparticles, chitosan, a biopolymer of glucosamine residues, has increased the attention for mucosal delivery system for vaccines. Chitosan is able to release high level of targeted antigen resulting in high long-lived antibody titers, which correlate with protection in case of influenza.\textsuperscript{21,22}

Hemokinin-1 (HK-1) as a molecular adjuvant can enhance circulating antibody responses when co injected with H9N2 whole inactivated influenza vaccine.\textsuperscript{16,23} Although the administration route elicits serum antibodies, it does not usually induce mucosal immune responses. Here, the scope of these initial studies was expanded to introduce a vaccine adjuvant/delivery platform capable of enhancing influenza humoral antibodies. The ability of chitosan nanoparticle-based HK-1 influenza vaccine candidate to provide protective immunity against influenza virus was evaluated by consideration of both adjuvant and delivery system.

Materials and Methods

\textit{Inactivated H9N2 influenza virus antigen preparation}

Madin-Darby Canine Kidney (MDCK) cell line (ATCCCL-34), the permissive cell for influenza virus replication, was maintained in DMEM medium (Sigma, Aldrich) supplemented with 10% FBS, 100 U/ml penicillin, and 100 μg/ml streptomycin at 37 °C in a humidified atmosphere of 5% CO\textsubscript{2}. Avian influenza H9N2 virus, A/Chicken/Iran/SS1/1998, was used in this study. Monolayers of the cells at a concentration of 1×10\textsuperscript{6} cells/ml were infected with the virus at a multiplicity of infection of 0.1 PFU/cell in the presence of supplemental trypsin. Following adsorption for 1 hr at 37 °C, the inoculum was removed and washed before DMEM was replaced. The cultures were incubated up to 48 hr post infection (hpi) and observed by inverted light microscopy for Cytopathic Effect (CPE). The virus was titrated by Hemagglutination Assay (HA) using 0.5% chicken RBCs.\textsuperscript{24} One HA unit (HAU) is equal to approximately 5 to 6 logs of the virus. The prepared H9N2 antigen was inactivated with a 0.02% final concentration of formalin (Merck) in PBS following incubation for 18 hr at 37 °C.

\textit{HK-1 adjuvant preparation}

HK-1 was prepared according to the previous study. Briefly, HK-1 sequences from Mus musculus were downloaded from the NCBI database and aligned by the Clustal W algorithm. The coding region was then amplified with the designed primer, HK-1F: 5’-GG ATCCCCCTGCCCCGTTCCTCCTGAT-3’ and HK-1R: 5’-CCATGGCTTCCCCATCAGACCCTAAT-3’ carrying the appropriate restriction enzyme sites and cloned into the pcDNA3.1 vector (Invitrogen, USA) between the BamHI and NcoI sites, respectively. Plasmid DNA was propagated in Escherichia coli (E. coli) and purified using EndoFree®Plasmid Mega Kit (Qiagen) according to the manufacturer’s instructions. The correct sequence of the cassette within pcDNA3.1 was confirmed by sequencing. Concentration of the purified plasmid was adjusted to 1 μg/μl.

Chitosan nanoparticle-based HK-1/influenza vaccine candidate preparation and characterization

At the first step, chitosan nanoparticles were made by ionic gelation method based on the ionic gelation of chitosan with sodium tripolyphosphate (TPP) anions (Nanoino Co, Iran). In brief, 0.2 g of the purified chitosan powder was dissolved in 100 ml 0.35% acetic acid solution and pH adjusted to 5.5 by 1 M NaOH after overnight incubation. 0.5% TPP was dropped to the chitosan solution under stirring condition for 30 min at room temperature. H9N2-loaded nanoparticles were formed by mixing 0.8 μl chitosan-TPP at concentration of 0.5 mg/ml by 0.1 ml H9N2 antigen containing 5 (log\textsubscript{2}) HA. The solution was incubated for 30 min at room temperature, then centrifuged at 12,000 rpm at 4 °C for 10 min. The amount of free antigen in the solution was determined by HA. The zeta potential (mV) value was measured by using ZetasizerNano ZS (Malvern, UK). Encapsulation efficiency and loading capacity of the nanoparticles were calculated as follows:

\[
\text{Encapsulation efficiency} = \frac{\text{Total HA} - \text{Free HA}}{\text{Total HA}} \times 100\%
\]

\[
\text{Loading capacity} = \frac{\text{Total HA} - \text{Free HA}}{1 \text{ mg chitosan nanoparticles dry weight}}
\]

By the same method HK-1/H9N2-loaded nanoparticles were made by adding 0.1 ml HK-1 (10 μg/ml) to the above solution. Stability of the prepared antigen-loaded and antigen/adjuvant-loaded nanoparticles during storage at 4 °C was evaluated by measuring the HA content using Hemagglutination (HA) assay. Briefly, 25 μl of the reagents were serially diluted 1:2 in PBS in a 96-well plate and incubated for 30 min at room temperature with 25 μl of chicken red blood cells (1% v/v). The
Influenza Nanovaccine

**Table 1.** Experimental groups of BALB/c mice used in the protective potential of HK-1/H9N2 nanovaccine (n=50)

<table>
<thead>
<tr>
<th>Group</th>
<th>Immunization scheme</th>
</tr>
</thead>
<tbody>
<tr>
<td>C1</td>
<td>Normal saline</td>
</tr>
<tr>
<td>C2</td>
<td>Inactivated H9N2 antigen</td>
</tr>
<tr>
<td>A1</td>
<td>Chitosan nanoparticle-based H9N2 antigen without HK-1 (prime)</td>
</tr>
<tr>
<td>A2</td>
<td>Chitosan nanoparticle-based H9N2 antigen without HK-1 (booster 14 days after prime)</td>
</tr>
<tr>
<td>B1</td>
<td>Chitosan nanoparticle-based HK-1/H9N2 antigen (prime)</td>
</tr>
<tr>
<td>B2</td>
<td>Chitosan nanoparticle-based HK-1/H9N2 antigen (booster 14 days after prime)</td>
</tr>
</tbody>
</table>

**Table 2.** The characteristics loaded H9N2 influenza antigen and hemokinin-1 as adjuvant on chitosan nanoparticles

<table>
<thead>
<tr>
<th>Particle size (nm)</th>
<th>Zeta potential (mV)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blank Ag/Adj loaded-nanoparticles</td>
<td>Blank Ag/Adj loaded-nanoparticles</td>
</tr>
<tr>
<td>254±21.7</td>
<td>43.6±1.2</td>
</tr>
<tr>
<td>338±22.6</td>
<td>47.7±0.9</td>
</tr>
</tbody>
</table>

The endpoint HA titer was defined as the reciprocal of the last highest dilution of reagent that caused complete agglutination of red blood cells. HA titers were re-determined once every two weeks to assess the loss of titer over a period of two months.

**Mice immunization trial**

Sixty healthy 6-8-week-old female BALB/c mice (Razi Vaccine & Serum Research Institute, Karaj) with 19 to 23 gr weight were used. The animals were equally distributed (Table 1) to the control groups (C1 and C2), which received normal saline and inactivated H9N2 antigen, respectively and four treatment groups as follows: groups A1 received chitosan nanoparticle-based H9N2 antigen without HK-1 and group A2 received booster 14 days after the primary vaccination. Group B1 received chitosan nanoparticle-based HK-1/H9N2 antigen and group B2 received the same vaccine 14 days after the primary vaccination. 5 μl of samples, regarding the groups, were dropped onto both eyes. Before administration, the vaccine was mixed with normal saline. Blood samples were taken from medial canthus of the eye up to 90 days post inoculation (pi) at 14 days intervals. Influenza virus antigen-specific antibody titers were determined by the endpoint titer dilution method using ELISA assay (IDEXX Influenza AAb test) according to the manufacture’s instruction.

**Statistical analysis**

The statistical analysis was performed with the use of SPSS version 15.0.1. p-value of <0.05 was considered statistically significant. The results were expressed as mean±SD for three independent experiments.

**Results**

The H9N2 virus was well replicated in MDCK cells in the presence of supplemental trypsin. The infected cell suspension was harvested following observation of CPE at 48 hpi (Figure 1). Virus HA titer was evaluated and found to be 3.70. The influenza virus antigen was further concentrated and formalin-inactivated. The coding region of HK-1 gene was amplified at 231 bp and electrophoresed on 1% agarose gel with the specific primers (Figure 2).

Chitosan microparticles were formed by ionic cross-linking between the positive charges of the amino groups of chitosan and the negative charges of TPP ions. Characteristics of the generated chitosan nanoparticle-based HK-1/H9N2 are shown in table 2. The larger size of the loaded nanoparticles than the chitosan-TPP may be due to the molecular weight of H9N2 virus. The zeta potential of the blank chitosan nanoparticles was +43.6, which displayed good protein adsorption. The addition of H9N2 inactivated antigen and HK-1 resulted in minor increase in the zeta potential due to the load of positively charged virus surface gly-
coprotein. Encapsulation efficiency of HK-1/H9N2 on chitosan nanoparticles was estimated 98.43% indicated that the antigen was sufficiently encapsulated. Stability of each nanoparticle formulations was assayed by maintaining the tested vials at 4°C for two months. The chitosan nanoparticle-based HK-1/H9N2 and chitosan nanoparticle-based H9N2 retained hemagglutinating activity up to the end of experiment compared to the inactivated H9N2 antigen. The HA titers (log2) of the nanoparticles remained 7 when tested after one month post incubation at 4°C, whereas the titer of H9N2 antigen dropped by 3 log2. At the end of experiment, no change on HA values of the nanoparticles was observed; while H9N2 antigen lost HA activity.

Efficacy of the generated chitosan nanoparticle-based HK-1/H9N2 vaccine candidate was evaluated in mice. The mice in vaccinated and control groups gained weight from 81.3 to 96.5%. All animals survived after vaccination indicating that the H9N2 antigen and the nanovaccine formulations were safe. The potential of HK-1 and chitosan nanoparticles was evaluated regarding whether they can enhance specific antibody production in response to H9N2 influenza vaccination. Chitosan nanoparticle-based H9N2 antigens with or without HK-1 induced humoral immunity in mice against H9N2 antigen, while loading H9N2 antigen and HK-1 onto the nanoparticles coated with chitosan carrier developed an improved immune response (Figure 3). The antibody production level was significantly higher (p<0.05) in groups B1 and B2 indicating chitosan nanoparticles loaded H9N2 antigen with HK-1 adjuvant induced immune responses against influenza; however, group B2 with twice vaccination showed a marginally higher antibody level. These results show that chitosan nanoparticle enhances immune responses in influenza immunization. Increased antibody levels in mice received chitosan nanoparticle-based HK-1/H9N2 antigen revealed that HK-1 adjuvant formulation conferred superior protection over the adjuvant-free formulation.

Discussion

Avian influenza is a significant global concern because of its potential to become the pandemic threat. New strategies should be taken into account in order to control the infection in human and bird populations. Studies with nanoparticle-based vaccine candidates evidenced their ability to stimulate mucosal as well as systemic immunity. In our study, it was shown that the chitosan as nano-antigen/adjuvant delivery system is effective to encapsulate viral antigen and induce immune response against influenza. The immunogenicity of the inactivated H9N2 antigen improved when HK-1 was used as an adjuvant.

Most current inactivated vaccines are administrated by intramuscular, subcutaneous, or intradermal injection. Although humoral immunity often provides good protection against influenza, mucosal vaccines may be necessary for induction of long-term protection. A variety of polycationic polymers and their derivatives were explored to stimulate immune system but they have not been an unqualified success, suggesting that both adjuvant and delivery system may need reconsideration. Some researches focus on the ability of chitosan nanoparticles as a novel adjuvant to induce specific antibody titer against influenza antigens. Sawaengsak et al. found that the chitosan nanoparticle encapsulated HA-split vaccine reduced markedly the influenza morbidity and also conferred 100% protective rate to the vaccinated mice against lethal influenza virus challenge. Khalili et al. also showed that the nanoparticles significantly enhanced the immunogenicity of inactivated H9N2 influenza virus in chickens in comparison to ISA70, an oil adjuvant. Similar results on chitosan nanoparticle advantages have been reported by other researchers. Taken together, loading influenza vaccines on chitosan nanoparticle and its derivatives can induce efficient immune responses against the viruses indicating the adjuvant property of these polysaccharides.

According to comparative analysis, it was found that H9N2 antigen plus HK-1 encapsulated into chi-
The improvement is dependent on the reversible antigen uptake by mucosal lymphoid tissues which im-
tation. The loaded chitosan nanoparticles easily release
actions or antigen only. This evidence supports chitosan
with chitosan-loaded H9N2 antigen and HK-1 formula-
peared to have a role in enhancing the systemic anti-
property 21,22,36. Here, it was found that chitosan ap-
role on induction of immunity indicated that the nano-
sion and antigen trans-muco sal presentation, and its
munity 34,35. The ability of chitosan for mucosal adhe-
phocytes, which can efficiently facilitate humoral im-
mentation is normally accompanied by the rapid class-
switching of the activated lymphocytes into IgA pro-
tion is normally accompanied by the rapid clasp
accuracy of chitosan are also added to the reversible bind-
action of chitosan with antigen/adjuvant that allows
antigen uptake and enhances antigenic trans-mucosal
delivery. The size, high positive charge, and hydropho-
licity of chitosan are also added to the reversible bind-
ing to make it a suitable delivery carrier 37.

In this study, an attempt was made to assess whether a booster vaccination could improve antibody responses
against influenza. Following the primary immunization,
antibody titers increased in mice either received chitosan nanoparticle-based HK-1/H9N2 or chitosan
nanoparticle-based H9N2 with a comparable level in control. In both treated groups, the booster vaccination
led to a rise in influenza specific antibody, which was not statistically significant between the prime and
boost in each group. These data suggest that a prime
immunization regimen conferred protection; however, formulation of influenza nanoparticle with HK-1 offers
a potential efficacy advantage.

**Conclusion**

It was proposed that eye drop vaccination of HK-
1/H9N2 nanoparticles is an effective and safe tool to
 induce protective immunity against influenza virus in-
fection. Other advantages of the vaccine candidate are
stability at 4°C which does not require low temperature
storage, good tolerability, non-toxic and easy admin-
istration. More studies need to be carried out to opti-
mize the adjuvanted-nanovaccine candidate for use in
commercial applications.

**Acknowledgement**

The authors thank Mohammad Reza Samiee and
Mosa Babaei for technical assistances. This work was
supported by Razi Vaccine and Serum Research Insti-
tute under grant No.2-2-18-94112.

**References**

1. Baigent SJ, McCauley JW. Influenza type A in humans,
mammals and birds: determinants of virus virulence, host
-range and interspecies transmission. Boessays 2003;25
(7):657-671.

2. Palese P, Garcia-Sastre A. Influenza vaccines: present

3. Cox RJ, Brokstad KA, Ogra P. Influenza virus: immunity
and vaccination strategies. Comparison of the immune
response to inactivated and live, attenuated influenza

genetic reassortants in H9N2 influenza A viruses and
their diverse pathogenicity to mice. Virol J 2011;8:505.

5. Lee YT, Kim KH, Ko EJ, Lee YN, Kim MC, Kwon YM,

6. Ben-Yedidia T, Arnon R. Epitope-based vaccine against

7. Hashem AM. Prospects of HA-based universal influenza


