Preliminary Study on Cost-Effective L-Tryptophan Production from Indole and L-Serine by E. coli Cells

Tahereh Sadeghiyan-Rizi 1, Jamshid Fooladi 2*, and Sima Sadrai 3

1. Department of Pharmaceutical Biotechnology, Faculty of Pharmacy, Isfahan University of Medical Sciences, Isfahan, Iran
2. National Laboratory of Industrial Microbiology, Department of Biology, Faculty of Science, Alzahra University, Tehran, Iran
3. Department of Pharmaceutics, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran

Abstract

Background: L-tryptophan is used widespread in the pharmaceutical industry. The majority of L-Trp production depends on microbial processes that produce L-tryptophan from indole and L-serine. These processes are very costly due to the costs of precursors, especially L-serine. Use of inexpensive substitutions as the L-serine source of L-tryptophan production enables us to reach a cost-effective process. In this paper, effect of Triton X-100 on L-Trp production and the ability to use Iranian cane molasses as inexpensive L-serine source was investigated.

Methods: Escherichia coli (E. coli) ATCC 11303 cells were grown in 10-L fermenter containing minimal medium supplemented with beet molasses as an inexpensive carbon source and indole as tryptophan synthase inducer. Whole cells of stationary phase were used as biocatalyst for L-Trp production. Triton X-100 addition to the production medium as indole reservoir was investigated. Then, cane molasses was used as L-Ser source in L-Trp production medium. Amount of L-Tryptophan and theoretical yield of L-Trp production was determined by HPLC and by a colorimetrically method on the basis of remaining indole assay, respectively.

Results: As a result, triton X-100 increased L-Trp production three times. Also, the result showed that 0.68 mM L-Tryptophan was produced in the presence of cane molasses at 37 °C for 8 hr.

Conclusion: This result showed that cane molasses of Qazvin sugar factory includes significant amounts of L-Ser that makes it a suitable substitution for L-Ser in L-Trp production. Therefore, it has the potential to be used for cost-effective L-Trp production in industrial scale.

Keywords: Molasses, PLP, Tryptophan synthase, Tryptophanase

Introduction

L-Trp is an important ingredient in medicines and animal feeds as an essential amino acid 1-3. Its use in infusion solution form as an antidepressant, sedative and in treatment of schizophrenia and alcoholism are some of the more important applications 4,5.

Several microbial processes used for L-Trp production include direct fermentation from carbohydrates or hydrocarbons 6,7, enzymatic reaction from L-Trp precursors 8 and bioconversion from L-Trp precursors 9. Many commercial processes employ Escherichia coli (E. coli) cells to convert indole and L-Ser to L-Trp by tryptophan synthase (TSase) 10. This process is very simple and a one-step reaction catalyzed by tryptophan synthase and when precursors are employed, the complete biosynthetic pathway of L-Trp isn’t necessary and the complicated mutations of microorganisms which control the regulatory mechanism are often not required 7. One disadvantage of this process is that precursors especially L-Ser and Pyridoxal Phosphate (PLP) are expensive. TSase is a PLP dependent enzyme. As established in our previous study, exogenous PLP isn’t needed in our reaction conditions because beet molasses contains relatively large amounts of sucrose, amino acids and vitamins such as pyridoxine and when used as the carbon source of culture medium not only accelerated the bacterial cells growth but also these cells gained a significant amount of PLP from the culture medium and when used as a biocatalyst in the
production medium, they didn’t require exogenous PLP as a cofactor of TSase unlike previous reports. This finding is important from an economical point of view. Therefore, in the present study, the amount of intracellular PLP was investigated. Another substrate, L-Ser, is very expensive because a racemic mixture is formed during its manufacture.

To solve this problem, new methods in both process design and cheap precursor substitution have been developed. Another disadvantage of this process is high concentration of indole inhibits TSase activity. To address this matter, organic solvents were used as reservoirs of indole in production medium.

The aim of this research was cost-effective production of L-Trp. Therefore, E. coli ATCC 11303 was cultured in the inexpensive medium and bacterial cells induced for TSase activity during growth by addition of indole to the culture medium harvested from growth medium were transferred to production medium as biocatalyst of L-Trp production from indole and L-Ser. The possibility of Triton X-100 use as an indole reservoir to prevent inhibition of TSase by indole and Qazvin cane molasses as a cheap L-Ser substitution for cost-effective L-Trp production was investigated.

Materials and Methods

Chemicals
Indole, L-serine, PLP, L-tryptophan, Triton X-100, TLC plates (60F254) and methanol were purchased from Merck (German), and all other chemicals were of analytical grade or the highest purity commercially available. Cane and beet molasses were obtained from Iranian sugar industries and cane and beet molasses composition is presented in table 1. Orumiyeh beet molasses was sterilized separately at 121°C for 15 min for preparation of culture medium and Qazvin cane molasses was passed through a 0.22 μm Millipore filter for preparation of reaction mixture.

Microorganism and culture conditions
E. coli ATCC 11303 was used in this research. For large scale production of biomass, a 500 ml culture, grown in complete medium containing glucose, 20 g/l; yeast extract, 25 g/l; (NH₄)₂SO₄, 0.5 g/l in 2 L Erlenmeyer flask on a rotary shaker, 180 rpm at 37°C for 10 hr, was inoculated into 10 L of a medium containing K₂HPO₄, 7 g/l; KH₂PO₄, 3 g/l; Na₃C₆H₅O₇.3H₂O, 0.5 g/l; MgSO₄.7H₂O, 0.1 g/l; Orumiyeh (northwest of Iran) beet molasses, 16.2 g/l and indole, 0.058 μmol/l. The bioreactor was a 10 L fermenter (type MS-F1-S-101) equipped with a draft tube and driven by a specially constructed propeller at 250 rpm and the pH of culture medium was adjusted automatically to pH 7.0±0.2 by titration with NaOH 10 N and HCl 1 N.

Determination of cell growth
The growth of microorganism was determined periodically by measuring the optical density at 620 nm every one hour and also 1 ml of culture medium was centrifuged at 8000 rpm at 4°C for 20 min and the harvested cell mass was dried at 105°C for 24 hr to a constant weight.

Tryptophan production
Biomass harvested from culture medium (3 g) was transferred to 500 ml Erlenmeyer flask containing production medium: 100 ml potassium phosphate buffer (0.1 M, pH=8), indole (0.05 g) and L-Ser (0.05 g), incubated on rotary shaker (180 rpm) at 37°C. After 4 hr of incubation, the production medium was centrifuged at 12000 rpm for 20 min in 4°C and the supernatant fluid was used for assay of produced L-Trp and remaining indole.

L-Trp was determined by HPLC in a Waters system equipped with isocratic pump and ultraviolet detector, by using an RP-18 column (MZ-analytical column, 4×100 mm) with 3 μm particle size and the mobile phase was phosphate buffer (0.05 M, pH=4.2): methanol (70:30). Injection volume, flow rate and UV absorption were 50 μl, 1.2 ml/min and 220 nm, respectively. For sample analysis, L-Trp was dissolved to a concentration of 1 mg/ml in purified water as stock solution. L-Trp concentrations of 5, 4, 2, 1, 0.5, 0.2, 0.1 and 0.05 μg/ml were prepared in purified water from stock solution and used for standard curve drawing. L-Trp concentration in the supernatant fluid of production medium was determined according to standard curve. AT 3000 Autochorm software was used for HPLC data analysis.

Consumed indol and the theoretical yield of L-Trp production was determined colorimetrically on the basis of remaining indole in the supernatant fluid of production medium at 490 nm by spectrophotometer. Briefly, indole concentrations of 0.1-1 μmol/ml were prepared with 96% ethanol and 0.5 ml of each concentration was transferred to the screw cap tube separately and 0.5 ml NaNO₂ and 2 ml Gly-HCl buffer (pH=2.6, 0.1 M) were added to it. All tubes were put to the beaker containing boiling water for 5 min. Following cooling down, the absorbance of them was measured at 490 nm and used for the standard curve. Indole concentration in the supernatant fluid of the production medium was determined according to the standard curve.

Determination of intracellular PLP
Bacterial cells (3 g) were sonicated at 100% amplitude for 6 min and centrifuged at 12000 rpm at 4°C for 20 min, and then the supernatant fluid was used for determination of the intracellular PLP by TLC. PLP concentration of 0.0001 g/ml was prepared in purified water as standard solution and kept at 4°C. TLC was performed on silica gel 60F254 plates with dioxane: deionized water 1:1 (v/v) as mobile phase. Before use, the plates were activated at 50°C for 30 min in an electrically controlled oven. After activation, the plates were cooled at room temperature and stored in a closed chamber at room temperature. Samples (30 μl) were applied to the plate by use of Hamilton syringe, 1 cm
above the lower edge of the plate. The spots were left
to dry completely and the plate was then developed by
the one-dimensional ascending technique in a rectangu-
lar chamber pre-equilibrated with the mobile phase for
15 min. After development, the plate was dried at 50°C
in an oven and the spots of PLP were visualized in 254
nm UV light 19,20.

Investigation of triton X-100 effect on L-Trp production
To avoid inhibition of TSase by indole, Triton X-
100, nonionic detergent, was used in the production
medium as reservoir of indole. This effect was investi-
gated by use of the 3 production media, one of them
without Triton X-100 (control sample), and 2 another
production media with 300 µl Triton X-100 and in one
of them Triton X-100 was added at first (sample 1) and
in another, it was added in 2 hr after production medi-
um incubation (sample 2).

Investigation of L-Serine content of Qazvin cane molasses
L-Ser in molasses has been investigated by TLC and
TLC scanning. 5 mg Qazvin cane molasses was diluted
in 10 ml distilled water, mixed with n-butanol (1:1),
vortexed, incubated at room temperature so that two
phases were formed then the organic phase (upper lay-
er) was harvested. Following evaporation, the residue
was mixed with 10 ml n-butanol and 50 µl of it was
loaded on the silica gel 60 F254 plate. The chromato-
gram was developed in rectangular glass chamber con-
taining mobile phase n-butanol/acetic acid/water (65:
13:22 V/V/V). The chromatogram was dried at 60°C
for 10 min and the ninhydrin reagent solution was
sprayed on the TLC and dried at 110°C for 5 min.
The identification was confirmed by Rf obtained by the
comparison of L-Ser standard. Then TLC scanning was
performed in the wavelength of 254 nm.

Investigation of L-Trp production by Qazvin cane molasses
Qazvin cane molasses (center of Iran) capability to
be used as L-Ser source of production medium was
investigated. L-Ser of production medium was dis-
placed with Qazvin cane molasses (5 g/100 ml of pro-
duction medium). Because other components except L-
Ser might exist in processed molasses and have an ef-
flect, reaction production medium was incubated for
longer time i.e. 10 hr and sampling was done at 4, 8
and 10 hr. Since cane molasses might contain L-Trp
and interfere with the results, L-Trp existing in produc-
tion medium was calculated before production medium
incubation (T0).

Statistical method
The experiments were carried out in triplicate. All
the results were analyzed statistically by one-way anal-
ysis of variance and Tukey test with 95% confidence
level using SPSS 20.0.

Results
Determination of cell growth
According to the results, E. coli cells reached to the
exponential phase of growth after 6 hr of incubation in

the fermenter and introduced to the stationary phase
(Figure 1). The bacterial biomass was recovered and
used as a biocatalyst in the L-Trp production medium.
The wet weight of harvested biomass was 105 g.

Investigation of triton X-100 effect on L-Trp production
As shown in figure 2, Triton X-100 is a suitable
indole reservoir and prevents inhibition of TSase by
indole. L-Trp production is increased 3 times by use of
Triton X-100.

Determination of intracellular PLP
As shown in the previous study, no exogenous PLP
was required as a cofactor of TSase due to the cellular
uptake of PLP from the molasses during culture in the
fermenter 11. This probability was investigated and
results showed that there was adequate amount of PLP
in the cells (Figure 3).

Investigation of L-Serine content of Qazvin cane molasses
Results obtained from scan TLC established that
there was about 0.025 g L-Ser in the 5 g of Qazvin
cane molasses (Figure 4).

Investigation of cane molasses usage instead of L-Ser
It was indicated that cane molasses of Qazvin sugar
factory includes significant amounts of L-Ser and 0.68
mM L-Trp was produced from it as the L-Ser source of
reaction (Table 2).

Discussion
As described above, L-Trp production from indole
and L-Ser was carried out using whole cells of E. coli
The cost-effective L-Trp production has gradually attracted the attention of the amino acid industry. The following conditions are very critical for cost-effective L-Trp production via biotransformation: 1) inexpensive medium for microorganism cultivation, 2) inexpensive substitutions for precursors, 3) a short reaction time.

In this research, beet molasses was used as an inexpensive carbon source of culture medium. Beet molasses contains relatively large amounts of sucrose, amino acids and vitamins such as pyridoxine. On this reason, in our study, use of beet molasses as carbon source of culture medium not only decreased the cost of microorganism cultivation but also bacterial cells gained significant amount of PLP from culture medium, as shown in figure 3, and when used as a biocatalyst in the production medium, exogenous PLP as cofactor of TSase was not required whereas in previous reports exogenous PLP had been needed and this finding is very important from the economic point of view.

The inhibition of TSase by indole is a major problem, so nonaqueous organic solvents or nonionic detergents are used as indole reservoirs. Results showed that L-Trp production is increased by use of Triton X-100 (Figure 2), presumably due to the increase in bacterial cells permeability and the gradual supply of indole to the reaction mixture of L-Trp production.

As the main result, L-Trp (0.68 mM) was produced from Qazvin cane molasses as L-Ser source unlike Fariman and Karaj cane molasses in our previous study. Cane molasses composition highly varies and depends on cane varieties, climate and processes of sugar production. However, it was observed that the use of cane molasses as L-Ser source of production medium showed a 32% decrease in L-Trp production in comparison to the time when L-Ser was used (Table 2). The result was expected beforehand because beet molasses contained considerable amounts of L-Trp and L-Trp is a strong inhibitor of L-Trp production. In the report presented by Mateus (1996), an initial L-Trp led to decrease in its initial production rate.

**Conclusion**

In this research it was proved that L-Trp production is increased by the use of Triton X-100 as indole reservoir in the production medium and also it was established Qazvin cane molasses includes significant amounts of L-Ser that makes it a suitable substitution for L-Ser in L-Trp production. This is the preliminary study on cost-effective L-Trp production by Iranian cane molasses and these results can be used in the industrial scale following further researches.

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References


