Inactivation of aprE Gene in *Bacillus subtilis* 168 by Homologous Recombination

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Abstract

**Background:** One of the most important producers of high quality industrial enzymes is the Gram-positive bacterium, *Bacillus subtilis* (*B. Subtilis*). One major limitation that hinders the wide application of *B. subtilis* is the secretion of high levels of extracellular proteases which degrade the secreted foreign proteins. In this study, homologous recombination technique was used to knock out its protease gene, aprE.

**Methods:** The internal segment of the pro-sequence of aprE gene of *B. subtilis* 168 with a length of 80 bps and its complementary sequence were synthesized and ligated into pUB110 at EcoR1 and XbaI restriction sites. Competent cells of *B. subtilis* 168 were prepared and transformed by electroporation using BioRad gene pulser as explained in the methods section. Transformants carrying the recombinant plasmid were selected for resistance to neomycin. The success of homologous recombination was checked by PCR amplification of the neo-mycin gene which was part of the vector and did not exist in the genome of *B. subtilis* 168. The protease activity was measured using the Protease Fluorescent Detection Kit based on the proteolytic hydrolysis of fluorescein isothiocyanate (FITC)–labeled casein-substrate.

**Results:** The results demonstrated that aprE gene would not be able to produce further active subtilisin E. The reduction of protease activity also confirmed the efficacy of the induced mutation in this gene.

**Conclusion:** It will therefore be a major challenge for future research to identify and modulate quality control systems of *B. subtilis* which limit the production of high quality protease-sensitive products such as lipase.

Keywords: aprE gene, *Bacillus subtilis*, Subtilisins

Introduction

*Bacillus subtilis* is a well-known Gram-positive soil bacterium that naturally produces and secretes high concentrations of proteins into the medium 1. The absence of an outer membrane in *B. subtilis* can simplify the protein secretion pathways and allow the organism to secrete high levels of extracellular proteins. In contrast to the Gram-negative bacterium *Escherichia coli*, *B. subtilis* is considered as a GRAS organism (Generally Recognized As Safe). Thus, scientists have widely used *B. subtilis* for commercial exploitation as a major "cell factory" for the secretion of heterologous proteins 2,3. As far as the capability of secreting extracellular enzymes directly into the culture medium is concerned, *B. subtilis* can potentially serve as an efficient expression host 4,5. The secreted foreign proteins usually remain in biologically active forms, and the downstream purification is greatly simplified 6,7.

One of the major limitations that hinder the
wide application of *B. subtilis* is the secretion of high levels of extracellular proteases which degrade the secreted foreign proteins. It is well established that *B. subtilis* has six extracellular proteases including neutral protease A, subtilisin (also known as alkaline protease), extracellular protease, metalloprotease, bacillopeptidase F, and neutral protease B. In view of the fact that the protease production is limited, protease deficient *B. subtilis* strains have been developed by genome engineering techniques. For example, *B. subtilis* WB800, deficient in eight extracellular proteases including neutral protease A, subtilisin (alkaline protease), extracellular proteases such as metalloprotease, bacillopeptidase F, and neutral protease B, was collected from the culture broth by centrifugation at 10,000 rpm. In order to improve the production of heterologous proteins, it can serve as an excellent host for the expression of heterologous proteins.

In order to improve the production of heterologous proteins in *B. subtilis* 168, it was decided to inactivate the aprE gene of this bacterium encoding one of the major extracellular alkaline serine proteases, Subtilisin E, by site directed mutation of this gene using homologous recombination techniques. For this purpose, the pro-sequence in the aprE gene was targeted. Subtilisin E is first made as pre-pro-subtilisin. This enzyme consists of a signal peptide for protein secretion (pre-sequence) and a peptide extension of 77 amino acid residues (pro-sequence) located between the signal peptide and the mature protease. Studies have indicated that the pro-sequence is essential for guiding appropriate folding of the enzymatically active conformation of Subtilisin E. Inactivating the pro-sequence of the aprE gene, therefore, should impair the production of an active subtilisin resulting in a genetically engineered strain deficient in one of the major extracellular proteases making an appropriate host for the expression of protease sensitive products.

**Materials and Methods**

**Bacterial strains, plasmids, and growth conditions**

The bacterial strain used in this study was *B. subtilis* strain 168 (DSMZ, Germany) containing the plasmid pUB110. The medium consisted of 7% maltose (w/v), 0.05% yeast extract (Difco), 4.6% dried corn steep liquor, 0.5% fish meal extract, 0.1% KH₂PO₄, 0.02% MgSO₄·7H₂O, and 15 µg/ml tetracycline (pH=6.8). The supernatant was collected from the culture broth by centrifugation at 10,000 g for 20 min and after that the pellet was resuspended in 1 mM EDTA, 1 mM PMSF, 1 mM DTT, and 5% glycerol in 50 mM potassium phosphate buffer (pH=7.4). Next, it was sonicated (model 7500; BIOMIC). The supernatants were then collected following centrifugation. Plasmid DNA from *B. subtilis* 168 was prepared as described by Sambrook et al.

**Insert preparation**

The internal segment of the pro-sequence of the aprE gene of *B. subtilis* 168 with a length of 80 bps (nucleotides 96 to 176 upstream from the initiation codon) and its complementary sequence were synthesized by Kawas Biotech Company (Iran). Restriction sites for the enzymes EcoRI and XbaI were designed on both ends of the single strands as outlined below:

5’-AATTCCagcagtacagaaaatatcattgtcgtcggatataaaacagacaatagtgcagcagctcagctcgcacgcaagaaagatgtT3’

and 5’-CTAGAacacatctctttctggccggaactcatggcactcat gtgcgttttaacctgcagatgttaccttcttctgtactgG3’

**Vector preparation**

Twenty five µl of PUB110 was cross digested with the enzymes EcoRI and XbaI in a total volume of 100 µl according to the manufacturer’s recommended conditions (Germany). Gel electrophoresis was also run to verify complete digestion of the plasmid PUB110.

**Ligation reaction and transformation**

The ligation reaction was carried out using an appropriate amount of vector and insert (a ratio of 3:1 insert to vector). One µl of 10× ligation buffer and 1 µl of Ligase were added to the mixture adjusting the final volume to 10 µl with ddH₂O. The ligation mixture was incubated overnight at 16 °C.

Competent cells of *B. subtilis* 168 were prepared and transformed by electroporation using BioRad gene pulser. Electroporative cells were prepared by diluting 2 ml of the overnight culture of *B. subtilis* 168 in 50 ml of fresh LB culture medium and grown at 37 °C to reach OD value of 0.3 at 600 nm. Then the cell suspension was cooled on ice for 10 min and harvested by centrifugation at 4 °C and 10000 g for 10 min. The cells were suspended in 5 ml ice-cold electroporation buffer and spun at 4 °C at 10000 g for 10 min. Finally, the cells were suspended in 1.5 ml of ice-cold electroporation solution and the competent cells were kept on ice until use. Ten µl (5 ng/µl) of the ligation mixture containing the recombinant plasmid pUB110 was added to 100 µl of electrocompetent cells and homogenized by gently mixing with pipette several times. The mixture was then transferred into a pre-chilled cuvette and an electric shock with a voltage of 1000 V and a time constant of 8.6 ms was applied. Following the electric shock, the cells were kept on ice for 10 min and then transferred to a pre-warmed LB plate and incubated for 24 h at 37 °C. The transformed colonies were picked with a sterile loop and grown on LB agar plate at 37° C.
shock, 2 ml of LB culture medium was immediately added to the electroporated cells followed by incubation for 1.5 hr at 37 °C.

Selection for neomycin resistance
Transformants carrying the recombinant plasmid were selected for resistance to neomycin.

Primer design for the neomycin gene and PCR amplification
To confirm the incidence of homologous recombination, PCR amplification of the neomycin gene was carried out. Neomycin gene was part of the plasmid pUB110 used as the vector and it did not exist in the genome of B. subtilis 168. The neomycin gene was amplified using the genomic DNA from transformed B. subtilis 168 as a template with a pair of primers, F (5’-TCGGAAAGTGACCAGA-3’) and R (5’-TTTGTGCCCTTATCGTAG-3’). PCR was conducted by subjecting the reaction mixture to an initial denaturation at 94 °C for 5 min, followed by 30 cycles of 94 °C for 1 min, 55 °C for 2 min, 72 °C for 3 min and a final extension step at 72 °C for 5 min.

Extracellular protease activity assay
To quantitatively compare the protease activity of wild and mutant strains deficient in active subtilisin E, the Protease Fluorescent Detection Kit (Sigma-Aldrich, Germany) based on the proteolytic hydrolysis of a fluorescein isothiocyanate (FITC)-labeled casein-substrate was used according to the manufacturer’s protocol. FITC-casein is native casein that has been labeled using fluorescein isothiocyanate. The UV absorbance of this heavily-labeled, intact protein substrate at 280 nm changed dramatically upon digestion by proteases, resulting in a measurable indication of proteolysis. Different concentrations of FITC-casein were also prepared by serial dilutions and their absorbance at 280 nm was measured to determine the concentration ranges across which the Beer-Lambert law was obeyed.

Results

Construction of recombinant plasmid, pUB110
To prepare the pUB110 for ligation, the plasmid was first subjected to restriction digestion with the enzymes EcoRI and XbaI. Digested plasmid was then dephosphorylated by alkaline phosphatase and recovered from the gel. The internal segment of pro-sequence in the aprE gene of B. subtilis was constructed with the same restriction enzyme sites (EcoRI and XbaI) as the vector. Agarose gel analysis of the pUB110 digested plasmid after dephosphorylation and EcoRI and XbaI digested insert is depicted in figure 1. Constructed insert and vector had the lengths of 80 and 4000 bps, respectively. An appropriate amount of prepared plasmid (pUB110) and insert were mixed for ligation reaction with T4 DNA ligase. Then recombinant plasmid was transformed into B. subtilis 168. Figure 1 shows the pUB110 plasmid digested with EcoRI and XbaI and also the 80 bp insert. The integrity of the plasmid (4 kb) of cut plasmid and the purchased insert both were verified.

Transformation of recombinant pUB110 plasmid into Bacillus subtilis 168
Screening the electroporated cells was done for antibiotic resistance and performed on nutrient agar plates supplemented with 5 and 10 mg/ml of neomycin 14. The colonies of transformed cells from the plasmid containing neomycin resistance gene successfully grew and formed a bacterial lawn on LB media as shown in figure 2. DNA extracted from the transformed cells was compared with non-transformed bacteria. Figure 3 shows the pattern of DNA extracted from the transformed and non-transformed bacteria.

PCR amplification of neomycin gene
To confirm the incidence of homologous recombination, PCR amplification of the neomycin gene was carried out. Neomycin gene was part of pUB110 and it did not exist in the genome of B. subtilis 168. Integration of this plasmid in the chromosome by double cross-over replaced the neomycin resistant gene in chromosomal DNA of B. subtilis.
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transformed *B. subtilis* 168. The neomycin gene was amplified using genomic DNA from transformed *B. subtilis* 168 as a template with a pair of designed primers. A sharp band in the 300 bps region (Figure 4) in the transformed cells proved the integration of the plasmid DNA into chromosomal DNA of transformed bacteria.

**Extracellular protease activity assay in wild and mutant strains**

Protease activity of samples was determined by measuring the residue of FITC-Casein in media. Activity measurement at the examined condition indicated that in mutant samples, the UV absorbance was increased in comparison with wild samples at 280 nm. These changes demonstrated that the aprE gene in *B. subtilis* 168 would not further be able to produce active subtilisin E and finally would decrease the protease activity.

**Discussion**

*B. subtilis* bacteria utilize especial systems that secrete proteins out of cells. Subtilisin E is one of typical examples which functions similarly. This extracellular alkaline serine protease encoded with aprE gene was produced at the end of exponential growth phase. Subtilisin E leads to degradation of other important proteins like lipase enzymes. It is known that if aprE gene is knocked out, *B. subtilis* will no longer be able to produce active subtilisin E and therefore, it can help to produce active proteins which are sensitive to proteases.

The aim of the present study was the inactivation of aprE gene using site directed mutation at pro-sequence region. For this purpose, the recombinant plasmid was constructed with mixing appropriate amount of insert and pUB110 plasmid as a vector. After transformation of recombinant plasmid into *B. subtilis* 168, screening results showed that electroporated cells can grow in supplemented media containing neomycin antibiotic. Comparison between transformed and non-transformed cells demonstrated that digestion, ligation and transformation were done successfully. The results of PCR amplification of neomycin gene further confirmed the homologous recombination. Finally, in accordance with our finding, the reduction of protease activity demonstrated that the
tation in aprE gene has indeed taken place at the right site.

**Conclusion**

As previously mentioned, *B. subtilis* is one of the most widely used bacterium for the production of industrial enzymes but its applications is limited due to its proteases. According to the results obtained in this study, homologous recombination techniques can help to inactivate protease encoding genes and improve production of other protease-sensitive proteins. It will therefore be a major challenge for future research to identify and modulate quality control systems of *B. subtilis* which limit the production of high quality protease-sensitive products such as lipase.

**References**